

The University of Southern Mississippi  
**The Aquila Digital Community**

---

EPA Study

Microbial Source Tracking

---

6-24-2009

## qPCR Method for Quantification of Human Polyomaviruses

Microbial Source Tracking

Follow this and additional works at: [https://aquila.usm.edu/mst\\_epastudy](https://aquila.usm.edu/mst_epastudy)

---

### Recommended Citation

Microbial Source Tracking, "qPCR Method for Quantification of Human Polyomaviruses" (2009). *EPA Study. 1.*  
[https://aquila.usm.edu/mst\\_epastudy/1](https://aquila.usm.edu/mst_epastudy/1)

---

This EPA Gulf of Mexico Alliance MST Study is brought to you for free and open access by the Microbial Source Tracking at The Aquila Digital Community. It has been accepted for inclusion in EPA Study by an authorized administrator of The Aquila Digital Community. For more information, please contact [aquilastaff@usm.edu](mailto:aquilastaff@usm.edu).

**Human Polyomavirus (HPyVs) Taqman PCR**  
**Published in Applied and Environmental Microbiology 2009**

***Reagents***

Reagent	Manufacturer	Cat. No.
TaqMan Universal PCR Master Mix, No AmpErase UNG	Applied Biosystems	4364343
MGBNFQ probe	Applied Biosystems	4316033

Oligonucleotide	Sequence	Length (bp)
Forward SM2	5'-AGT CTT TAG GGT CTT CTA CCT TT-3'	23
Reverse P6	5'-GGT GCC AAC CTA TGG AAC AG-3'	20
Probe KGJ3	5'-(FAM)-TCA TCA CTG GCA AAC AT-(MGBNFQ)-3'	17

***Standard curve***

Plasmids containing either the BK virus (ATCC VR-837) or JC virus (ATCC VR-1583) insert are used for the standard curve. Insert copy numbers are estimated by multiplying the average DNA concentration by Avogadro's number then dividing by the product of the insert length and average weight of a base pair (1). To produce a standard curve, the recombinant plasmid DNA is serially diluted in nuclease-free reagent grade water to a final concentration ranging from  $10^2$  to  $10^6$  gene copies• $\mu\text{l}^{-1}$ .

***qPCR reaction master mix***

Reagent	Volume ( $\mu\text{l}$ )	Final concentrations
Taqman Master Mix	25.0	n/a
SM2 (10 $\mu\text{M}$ )	2.5	0.5 $\mu\text{M}$
P6 (10 $\mu\text{M}$ )	2.5	0.5 $\mu\text{M}$
KGJ3 (10 $\mu\text{M}$ )	2.0	0.4 $\mu\text{M}$
Sterile water	13.0	n/a
Template	5.0	n/a
Total	50.0	n/a

***qPCR reaction conditions***

Amplification was performed in the 7500 Real-Time PCR System (Applied Biosystems, Foster City, CA).

	Temperature (°C)	Time	
DNA polymerase activation	95	10 min	
Denaturing	95	15 sec	
Anneal	55	20 sec	
Extension	60	60 sec	45 cycles

In general, Applied Biosystems default settings for the threshold cycle (Ct) are used for data analysis. However, amplification curves should be examined to ensure the automatic Ct is appropriate for the samples processed.

***Questions or concerns***

Contact:

Shannon McQuaig  
mcquacker@aol.com

Jody Harwood  
vharwood@cas.usf.edu

**REFERENCES**

1. **Yun, J. J., L. E. Heisler, Hwang, II, O. Wilkins, S. K. Lau, M. Hyrcza, B. Jayabalasingham, J. Jin, J. McLaurin, M. S. Tsao, and S. D. Der.** 2006. Genomic DNA functions as a universal external standard in quantitative real-time PCR. *Nucleic Acids Res* **34**:e85.
2. **McQuaig, S. M., T. M. Scott, J. O. Lukasik, J. H. Paul & V. J. Harwood.** 2009. Quantification of human polyomaviruses JC Virus and BK Virus by TaqMan quantitative PCR and comparison to other water quality indicators in water and fecal samples. *Appl Environ Microbiol* **75**: 3379-3388.