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CHANGES IN THE MORPHOLOGY OF WIDGEON GRASS (RUPPIA MARITIMA) WITH THE ONSET OF REPRODUCTION AND IMPACTS ON FISH ASSEMBLAGES AT THE CHANDELEUR ISLANDS, LA

Matthew Byrnes

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CHANGES IN THE MORPHOLOGY OF WIDGEON GRASS (RUPPIA MARITIMA)

WITH THE ONSET OF REPRODUCTION AND IMPACTS ON FISH

ASSEMBLAGES AT THE CHANDELEUR ISLANDS, LA

by

Matthew Byrnes

A Thesis Submitted to the Graduate School, the College of Arts and Sciences and the School of Ocean Science and Engineering at The University of Southern Mississippi in Partial Fulfillment of the Requirements for the Degree of Master of Science

Approved by:

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ABSTRACT

Seagrasses are important submerged coastal habitats that support nearshore communities. *Ruppia maritima* (widgeon grass) is a widespread species whose leaves branch extensively when reproductive. The goal of this study was to understand use of reproductive and non-reproductive *R. maritima* by fishes, recognizing the morphological change undergone by the plant when flowering. During the peak reproductive season in August and September 2021, *R. maritima* meadows at the Chandeleur Islands, LA were sampled to describe the spatial distribution and morphology of reproductive plants and investigate habitat use by fish assemblages. We assessed spatial trends in *R. maritima* presence and occurrence of reproduction and evaluated differences in shoot morphology using the metrics of longest leaf length, surface area, number of branches, and root to shoot ratios. We calculated density, Shannon diversity, and species richness to describe fish assemblages in reproductive and non-reproductive meadows. Additionally, general additive models were used to predict drivers of fish assemblage metrics. Results indicate that *R. maritima* was distributed along the entire length of North Chandeleur Island, but reproductive plants were located in the central, protected portion of the island. Reproductive plants were more morphologically complex than non-reproductive plants, but this did not drive fish assemblages. Rather, fish abundance was related to *R. maritima* biomass. This study provides information on patterns and drivers of habitat use by fish in *R. maritima*-dominated ecosystems that can be used to inform management and restoration.

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DEDICATION

To my family and friends who never leave my side through all the chaos of life. Also, I would be remiss if I failed to acknowledge my dog Cash who has been offering critical reviews as I write this document.

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1.1 Introduction

1.1.1 Seagrasses

Seagrasses are submerged angiosperms that inhabit marine environments, and include over 60 species across 13 genera (Larkum et al., 2006). The majority of seagrass genera occur at temperate and tropical latitudes in the northern and southern hemispheres (Larkum et al., 2006), and species' distributions are determined by taxon-specific light, nutrient, salinity, temperature, pH, and substrate requirements (Hemminga & Duarte, 2000; Larkum et al., 2006; McKenzie et al., 2016).

Seagrass expansion can occur through asexual or sexual processes. Clonal reproduction (asexual propagation) through fragmentation or rhizome extension is considered the primary mechanism for meadow formation and maintenance (Olesen et al., 2004); however, seagrasses also have the capacity for sexual reproduction, with species either being monoecious (i.e., both male and female organs on a single plant) or dioecious (i.e., separate male and female plants) (Larkum et al., 2006). A majority of seagrasses (42 species) produce dormant seeds that create seed reserves in the sediment near the parent plants that provide a buffer against disturbance (Darnell et al., 2021); in highly disturbed environments, dormant seeds are essential to the recolonization process (Larkum et al., 2006; Olesen et al., 2004).

1.1.2 Seagrass as Habitat

Seagrasses are essential submerged coastal habitats that support nearshore communities around the globe. These habitats often support faunal communities that are highly diverse and productive (Castillo-Rivera et al., 2002) and include valuable recreationally and commercially fished species (Bertelli & Unsworth, 2014; Nordlund et al., 2018). Seagrasses provide a variety of ecosystem functions specifically to fishes including shelter from predators and productive feeding grounds (Heck et al., 2003; Nordlund et al., 2018; Vaslet et al., 2012). Animals often rely on seagrass habitats for particular life stages, most importantly as nursery habitat for juveniles. McDevitt-Irwin et al. (2006) conducted a meta-analysis of 51 studies and found that seagrasses tended to support greater abundances of juveniles with higher survival rates compared to marsh, reef, mangrove, and other benthic habitats. Juveniles of many species utilize the abundant food and resources within the seagrasses to achieve faster growth rates prior to moving to more competitive adult habitats (McDevitt-Irwin et al., 2016).

1.1.3 Drivers of Fish Assemblage Composition

Drivers of site-specific fish assemblages in seagrass habitats are widespread and include local environmental conditions as well as the species and morphology of the seagrasses present, and at a broader scale, climate patterns. In a review of fish assemblages of northern Florida sub-tropical seagrasses, Fodrie et al. (2010) reported that more tropical fishes were present as sea surface temperatures increased, indicating "tropicalization". Similar observations have been reported along the Texas coast, with increases in temperature and sea level corresponding to an increased presence of tropical species (Fujiwara et al., 2019). Hyndes et al. (2016) hypothesizes that these changes will initially cause an increase in biodiversity until a new biological equilibrium develops. Climatic changes can also affect abiotic conditions (e.g., dissolved oxygen) that may impact seagrass habitat suitability for certain species (Francis, 2013).

Distinct fish assemblages among seagrass species can be linked to differences in plant morphology (Nakaoka, 2005; Rotherham & West, 2002). Morphological characteristics (e.g., leaf number and length and shoot density) create microhabitats that may be favored by certain species because of the food, shelter, and/or other ecological benefits they provide. Hyndes et al. (1996), for example, reported that whiting species (Sillaginidae) in southwestern Australia showed preference for *Zostera* spp. because the less dense canopy allows for easier movement when compared to the other local seagrasses *Posidonia australis* and *P. sinuosa*.

1.1.4 Role of Seagrass Complexity

Plant complexity impacts the function of seagrasses as hunting grounds, shelter, and juvenile nursery habitat (Larkum et al., 2006). When discussing complexity, it is often categorized by spatial scales starting at the seascape level then moving to finer resolutions within patches and at the individual plant level (Horinouchi, 2007; Hyndes et al., 2003; Skilleter et al., 2017). Seascape complexity involves the arrangement of habitats within systems. This can include systems which are characterized by monotypic seagrass beds, mixed species beds, the spatial arrangement and/or sized of meadows, or a combination of nearshore habitats such as seagrass, coral reef, oyster reef, and mangroves (Nagelkerken et al., 2002; Olds et al., 2012; Staveley et al., 2017, 2020). Fish species have variable responses to increased complexity, depending on their on their size and life history. Cruising predators, for example, benefit from meadow fragmentation as more edge habitat is created, which increases their hunting efficiency (Mahoney et al., 2018). Despite this, less complex seagrass seascapes generally have more diverse fish assemblages (Staveley et al., 2020), with many species benefitting from the shelter and

resources provided by large continuous seagrass meadows (Hovel et al., 2002; Williams et al., 2016). Individual seagrass patches can vary in density and substrate features (e.g., the presence of rocks or corals) (Horinouchi, 2007). Patches which are more dense may provide more niche spaces for predator avoidance and food when compared to less dense patches that are easier for predators to move through (Ambo-Rappe, 2016), and substrate features may increase fish diversity and abundance by creating unique microhabitats and increased shelter for smaller prey species (Cuadros et al., 2017). At the finest resolution, complexity is driven by individual plant morphology (e.g., leaf length, width, and number) (Larkum et al., 2006). Micheli et al. (2008) reported that beds of *Z. marina* with greater leaf surface area supported higher fish abundances when compared to shoal grass (*Halodule wrightii*) that has a lower leaf surface area. Different fish species also exhibit preferential distribution during certain life stages, with smaller individuals frequently associating more closely with morphologically complex seagrass species (Middleton et al., 1984).

1.1.5 Widgeon grass (*Ruppia maritima***)**

Ruppia maritima is a widely distributed seagrass with a wide tolerance to environmental conditions that allows it to thrive in waters ranging from fresh to hypersaline (Hartog & Kuo, 2006) in both the northern and southern hemispheres from tropical to temperate latitudes (Orth & Moore, 1988; Reyes & Merino, 1991). *R. maritima* is an early successional species that is often the first species to colonize an area following a disturbance event (Cho et al., 2009). This is due to high rates of shoot turnover and the species' reliance on sexual reproduction that creates a persistent seed bank buried in the sediment (Kilminster et al., 2015). In some areas, *R. maritima* is

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perennial, whereas in more ephemeral areas, plants are predominantly annual (Malea et al., 2004). *Ruppia maritima* is monecious, having both male and female reproductive structures on a single plant. When reproductive, the morphology of *R. maritima* shoots change dramatically; whereas non-reproductive *R. maritima* shoots have several narrow (1–2 mm) leaves ranging from 5–20 cm in length (Kantrud, 1997; Larkum et al., 2006), reproductive shoots branch extensively and can reach a length of 2.5 meters (Larkum et al., 2006).

Figure 1.1 *Comparison of non-reproductive and reproductive* R. maritima *shoots: (A) non-reproductive* R. maritima *(Radloff et al., 2013) ; (B) reproductive* R. maritima *(Kantrud, 1991)*

R. maritima plants at temperate latitudes undergo one annual reproductive cycle, typically in the summer months (Bigley & Harrison, 1986), while those found in more tropical regions usually complete two reproductive cycles (Orth & Moore, 1988; Pulich, 1985). Cho and Poirrier (2005) reported that *R. maritima* in Lake Pontchartrain, LA, flowers in Spring (March to May) and again in late Summer/early Fall (August to October).

The change in plant growth form with the onset of reproduction increases structural complexity and has the potential to impact habitat use by fish and invertebrates,

as the reproductive shoots may indirectly provide increased opportunities for food and shelter through the creation of additional microhabitats. It is critical to understand patterns in the distribution of reproductive plants as a predictor of the habitat value of *R. maritima*, especially in areas experiencing changes in seagrass distribution and shifts in species composition.

1.1.6 Chandeleur Islands, LA

The Chandeleur Islands, LA, a chain of barrier islands in the northern Gulf of Mexico, represent an area of shifting seagrass species distribution and composition. The island chain stretches 72 km, with the northern end located 35 km south of Biloxi, MS and the southern end lying in an arch 25 km northeast of Venice, LA (Poirrier & Handley, 2007).

Figure 1.2 *Map of the northern Gulf of Mexico with the North Chandeleur Island, LA study site bounded by the red box*

Much of the land mass of the Chandeleur Islands is in the northern islands, with North Chandeleur Island being the northernmost and largest island. The islands are suffering chronic land loss due to a lack of sediment input, hurricane damage, and rising sea levels (Moore et al., 2014). Additionally, the *Deepwater Horizon* oil spill in 2010 impacted sediment and vegetation along the islands with both showing increased levels of total polycyclic aromatic hydrocarbons (Kenworthy et al., 2017).

The back barrier shelf to the west of the islands is protected from wave action which allows for the proliferation of extensive seagrass meadows (Ellinwood, 2008). There are five species of seagrass that grow along the leeward protected side of the islands, with the most common species being *R. maritima*, shoal grass (*Halodule wrightii*), and turtlegrass (*Thalassia testudinum*) (Kenworthy et al., 2017). Manatee grass (*Syringodium, filiforme*) and star grass (*Halophila engelmannii*) are also present, but are not as abundant (Kenworthy et al., 2017). Seagrass meadows at the Chandeleur Islands represent the only mixed meadows of these five species along 1,000 km of coastline from Perdido Key in Florida to the Texas Coastal Bend (Darnell et al., 2017). Seagrass cover has been decreasing along the island chain with 15,758 acres in 1969 to only 2,614 acres in 2011 (Handley & Lockwood, 2020; Pham et al., 2014). The observed decline is linked to a reduction of shallow protected areas through the combined processes of changes in island geomorphology and storm-related land loss (Darnell et al., 2017). For example, in the aftermath of Hurricane Katrina, the Chandeleur Islands lost approximately 70% of their land mass and 20% of the seagrass cover (Bethel & Martinez, 2008). In the wake of these disturbances, there has also been an observed shift in seagrass species composition, with increase in cover of *R. maritima* and decrease in cover of other species such as *T.*

testudinum (Kenworthy et al., 2017). Large seed banks and rapid growth are characteristics that allow *R. maritima* to thrive after large disturbances and out-compete other seagrass species (Cho et al., 2009; Poirrier & Handley, 2007). As disturbances increase in frequency, as projected with impacts of climate change (Collins et al., 2019), it is likely that *R. maritima* will continue to increase in cover at the Chandeleur Islands, with unknown impacts to the seagrass-associated communities.

1.1.7 Objective

The goal of this study was to understand the distribution of *R. maritima* and describe the morphology of reproductive plants at the Chandeleur Islands. Then, to quantify the impacts of reproductive and non-reproductive *R. maritima* on habitat association by fishes, testing the following null hypotheses:

> H01: Reproductive *R. maritima* plants will be homogenously distributed across the Chandeleur Islands

H02: Fish assemblages will not differ between reproductive and nonreproductive *R. maritima.*

1.2 Methods

1.2.1 Vegetation Sample Collection

To describe the distribution and morphology of reproductive and nonreproductive *R. maritima* at the Chandeleur Islands, *R. maritima* was surveyed across its extent at the islands during late summer (August/September) 2021. Prior to sampling, sites with historic presence of reproductive and non-reproductive *R. maritima* were identified using seagrass monitoring data collected in previous years (K. Darnell, unpub. data). Those sites where *R. maritima* previously occurred ($n = 30$) were targeted for this

study, while also ensuring sites were distributed across the entire area of seagrass occurrence at the Chandeleur Islands.

At each sampling site, a YSI handheld meter (Pro 2030, YSI Inc., Yellow Springs, OH) was used to measure salinity, dissolved oxygen (mg/L) , and temperature (C°) ; water depth (cm) was measured using a pole marked in 5 cm increments; and light irradiance at the surface and at ~58–cm depth (μ m photons m⁻² s⁻¹) was measured using two, four pi (spherical quantum) sensors and a data logger (LI-1500, LI-Cor., Lincoln, NE). Light attenuation coefficients (k_d) were calculated as the inverse natural log of the deeper sensor value divided by the surface value, then divided by 0.58-m (linear distance between sensors). At each site, seagrass percent cover by species and bare sediment were quantified in three quadrats (50-cm \times 50-cm), and the percent cover of reproductive and non-reproductive *R. maritima* was estimated. One seagrass core (9-cm diameter \times 15-cm depth) was collected within *R. maritima* in each quadrat to quantify shoot density and biomass of reproductive and non-reproductive shoots. Quadrats were only collected from sites with seagrass present and cores were only collected from quadrats with seagrass cover. Cores were placed in plastic bags and transported to the Gulf Coast Research Lab (GCRL) where they were frozen until processing. Five reproductive and five nonreproductive plants were retained from the site for image analysis, with care taken to collect all aboveground (leaf and flower) and belowground (root and rhizome) tissue for each plant.

1.2.2 Fish Sample Collection

Fish were sampled at the Chandeleur Islands, LA, during a two-week period from September 6–11, 2021, at the height of the *R. maritima* reproductive season in this area.

This sampling time frame captures the greatest contrast in plant complexity between reproductive and non-reproductive plants. Fish were sampled using a throw trap at a subset of sites surveyed for differences in plant distribution and morphology. Throw trapping offers a targeted approach to sampling patchy habitat that is not possible with other gears such as benthic sled or trawl which cover larger areas (Camp et al., 2011; Jordan et al., 1997). The throw trap consisted of a 1-m \times 1-m \times 0.6-m aluminum frame. Nylon mesh (3.175 mm) was sewn onto the frame sides as well as extended above the frame and attached to floats to extend the trap height to 1.5 m. After trap deployment, all vegetation was removed and retained for processing. A bar seine was used to remove organisms from within the trap. The bar seine (90-cm \times 50-cm) consisted of a PVC frame with handles and 3.175-mm mesh stretched between. The pattern for seining organisms was modeled from Shakeri et al. (2020) and involved three sweeps from each of the trap sides, with sweeps continuing until three consecutive sweeps yielded no additional fish. Collected fish were placed into plastic bags and stored on ice until returned to GCRL, where they were frozen until processing.

Sampling was conducted in *R. maritima* patches across a range of reproductive plant coverage. Patches with desired characteristics (e.g., target cover of reproductive and non-reproductive plants) were identified from those sampled in the survey for plant distribution and morphology and marked with PVC poles prior to sampling. Sampling depth was limited <1.5-m due to the height of the throw trap, and sites were separated by at least 15-m to minimize effects of disturbance. Thirty sites were selected to span the range of reproductive cover values.

1.2.3 Vegetation Sample Processing

Seagrass cores were rinsed over a 500-um sieve and plants were separated by species. The number of shoots of each species was counted, but only *R. maritima* plants were retained. The number of shoots, the reproductive status of each shoot, and the number of branching nodes per plant were recorded. Epiphytes were removed from leaves by gently scraping both sides of each leaf with a razor blade; epiphytes were then placed in a drying oven for 48 h at 60° C to obtain a dry weight to the nearest thousandth of a gram. Shoots were classified as reproductive or non-reproductive and separated. Reproductive shoots were defined as those with branching leaves and/or the presence of inflorescences. The aboveground and belowground tissues were then separated, and the aboveground and belowground tissues for each shoot type (reproductive or nonreproductive) were grouped and placed in a drying oven for 48 h at 60° C, after which they were weighed to the nearest thousandth of a gram to obtain a dry weight. Dried biomass was used to calculate a core-level root to shoot ratio (RSR), which is a useful proxy for plant condition and productivity (Hitchcock et al., 2017). Measurements from the 9-cm diameter core were then extrapolated to obtain bed characteristics per square meter.

Digital images (600 dpi) were taken of the handpick plants using a high-resolution flatbed scanner (Epson WF-3640). Prior to scanning, epiphytes were removed from each shoot and the aboveground and belowground tissues were separated. The aboveground biomass was scanned to produce a JPEG format image. The total leaf area $\text{(mm}^2)$ was calculated using ImageJ (Version 1.53) with a threshold processing procedure (Easlon $\&$ Bloom, 2014). Additionally, the number of branching nodes and shoot length (mm) were

recorded from the image. Total leaf surface area, number of branching nodes, and shoot length were used to quantify overall plant complexity. Each individual shoot's epiphyte, and aboveground and belowground biomass was dried separately in a drying oven for at least 48 h at 60 \degree C, after which they were weighed to the nearest thousandth of a gram to obtain dry weights (g). This information was used to calculate an individual plant RSR.

Seagrass collected from the throw trap was returned to GCRL and sorted for nekton, then the plant biomass was spun for 90 seconds using an industrial sized salad spinner to remove excess water and sorted and weighed by seagrass species to obtain a species-specific wet weight (g) . Individual seagrass species wet weights were then combined to get a total wet weight. Finally, a random subset of reproductive and nonreproductive *R. maritima* plants (at least 3 reproductive and 3 non-reproductive plants per throw trap sample) were imaged and dried as described above to quantify total leaf area and biomass.

1.2.4 Fish Sample Processing

Fish were identified to species and the number of individuals of each species was counted. All individuals were measured for standard length (SL mm) (except for Dwarf Seahorses, which were only measured for total length), total length (TL mm), and weighed (g).

1.2.5 Vegetation Data Analysis

Sampling sites were mapped using QGIS (version 3.18.1) to display the spatial distribution of *R. maritima* along the Chandeleur Islands. Sites with reproductive *R. maritima* were also mapped to show the spatial distribution of reproductive plants. Each map was then evaluated for qualitative trends such as spatial clustering. The mean and

standard error were calculated for abiotic parameters across all sites to characterize the distribution of site conditions across the study area.

Several metrics were selected to compare plant morphology between reproductive and non-reproductive shoots. These metrics included surface area, shoot length, leaf number, and RSR. Metrics were compared using unpaired two-sample t-tests (alpha = 0.05). Prior to testing each metric, the data were evaluated for violations of the parametric testing assumptions. If assumptions were violated the sample was bootstrapped with resampling and the test statistic calculated. This was repeated for 10,000 permutations and the reported test statistic was represented by the mean of bootstrapped test statistic distribution.

1.2.6 Fish Data Analysis

Fish assemblages were compared across sites using density, Shannon diversity, and species richness. Density was calculated as the number of individuals per square meter. Shannon diversity (H') was calculated using standard methods (Freeman et al., 1984). Species richness was calculated as the number of unique species.

General Additive Models (GAM) were formulated to evaluate drivers of fish assemblage metrics (Shannon diversity, species richness, and density) with three potential predictor variables: water depth (cm), biomass of vegetation recovered from throw trap (g), and the proportion of reproductive *R. maritima* in the throw trap. Prior to running the GAMs, the predictor variables were plotted against each other to identify any significant relationships. Biomass of vegetation recovered from throw trap and the proportion of reproductive *R. maritima* in the throw trap were found to co-vary; the final GAM formulations only included biomass of vegetation recovered from throw trap and depth as

predictor variables. Poisson distributions were used to model species richness and density, while a gaussian distribution was used for modeling Shannon diversity.

1.3 Results

1.3.1 Reproductive Plant Spatial Distribution

Twenty-seven sites were sampled from August 5 to September 3, 2021 spanning the length of seagrass distribution along the Chandeleur Islands (N 30.0224°, W- 88.86616° to N 29.77237°, W -88.88668°)(Figure 1.1). Abiotic parameters were within the known ranges for seagrass meadows at the Chandeleur Islands (Darnell et al., 2017; Hayes, 2021) (Table 1.1).

Figure 1.3 *Sites sampled for* R. maritima *reproductive status during August and September 2021. Each point represents an individual sample site, sampled with three 0.25m² quadrats.*

Abiotic Metric	Mean \pm SE
Depth (cm)	97.2 ± 7.60
Temperature $(^{\circ}C)$	30.8 ± 0.44
Dissolved Oxygen $(mg L^{-1})$	7.82 ± 0.52
Salinity (ppt)	18.97 ± 0.67
Light Attenuation Coefficient m^{-1}	1.08 ± 0.06

Table 1.1 *Abiotic measurements for 27 sites sampled for the presence and distribution of reproductive* R. maritima

Of the sites sampled, 24 (88.9%) had *R. maritima* present, and 17 of the sites (63.0%) had reproductive plants (Figure 1.2). Among the 72 quadrats sampled across the 27 sites, mean (\pm SE) percent cover of non-reproductive *R. maritima* was 44.5 ± 3.5 , mean percent cover of reproductive *R. maritima* 21.5 ± 3.2 , and mean canopy height was 216.8 ± 6.3 mm. Sites with reproductive plants showed a latitudinal central tendency along the islands (Figure 1.2).

A total of 65 cores were collected across the 24 sites with *R. maritima* present. The mean (\pm SE) overall shoot density was 3,866.71 \pm 293.01 shoots per m², mean nonreproductive *R. maritima* shoot density was $3,843.13 \pm 299.37$ shoots per m², mean reproductive *R. maritima* shoot density was 895.94 ± 194.22 shoots per m², and mean core RSR was 1.097 ± 0.111 . Within the cores, the density of reproductive shoots was significantly greater than the density of non-reproductive shoots $(t = 8.2589, P < 0.05)$. Out of the 65 cores, only 19 cores had measurable epiphyte cover $(0.068 \pm 0.041$ g for all cores combined). Mean epiphyte biomass for cores with only non-reproductive shoots

 $(n=10)$ was 0.098 ± 0.075 g and epiphyte biomass for cores containing both nonreproductive and reproductive shoots (n = 9) was 0.035 ± 0.026 g.

Figure 1.4 *Presence of reproductive* R. maritima *at the Chandeleur Islands, LA during August and September 2021. White symbols indicate sampling stations with reproductive* R. maritima *present, while red symbols indicate stations where only non-reproductive* R. maritima *was present. Black symbols indicate stations where* R. maritima *was completely absent.*

1.3.2 Plant Morphology

A total of 74 reproductive and 112 non-reproductive plants were hand-collected to assess differences in plant morphology. Reproductive plants had significantly longer leaves (t = 7.054, p<0.05), greater total surface area (t = 11.509, p<0.05), and a greater number of leaves (t = 15.005, p<0.05), while non-reproductive plants had a higher RSR (t $= 10.440$, p < 0.05), indicating a larger contribution of belowground biomass (Table 1.2).

* Value significantly greater $(p<0.05)$ than the compared value

Table 1.2 *Mean morphological metrics for individual reproductive and non-reproductive hand-collected* R. maritima *plants.*

1.3.3 Fish Assemblages

A total of 22 sites were sampled to investigate habitat use by fishes. Sites were predominant located the center of the island where the most reproductive *R. maritima* plants were observed. The mean $(\pm \text{ SE})$ depth was 71.6 ± 2.8 cm, mean salinity was 19.46 \pm 0.37 ppt, mean temperature was 28.81 \pm 8.64 °C, and mean dissolved oxygen was 8.15 ± 0.02 mg/L. Mean (\pm SE) total percent cover seagrass was 69.1 \pm 2.9 %, mean percent cover non-reproductive *R. maritima* was 38.9 ± 5.2 %, mean percent cover reproductive *R. maritima* was 30.2 ± 4.3 %, mean total *R. maritima* wet biomass was $(371.58 \pm 27.61$ g).

Figure 1.5 *Sites sampled for fish assemblages within* R. maritima*. Each white circle represents a general sampling area, with the number inside the white circle indicating the number of samples taken in that area.*

Across all sites, a total of 224 individual fish were collected across 15 species.

Mean (\pm SE) fish abundance was 10.2 ± 1.7 per m², mean species richness was 3.4 ± 0.2 , and mean Shannon diversity was 0.97 ± 0.07 (Table 1.4). The five most abundant species accounted for 86.6% of all individuals collected and included Darter Goby (n=107), Blackcheek Tonguefish (n=33), Gulf Pipefish (n=23), Rough Silverside $(n=20)$, and Code Goby $(n=11)$. The mean weights, total lengths, and standard lengths for all species are listed in Table 1.3**.**

Species	Count	Mean Standard Length (mm) \pm SЕ	Mean Total Length (mm) \pm SE	Mean Individual Wet Weight (g) \pm SЕ
Darter Goby	107	24.05 ± 0.66	30.81 ± 0.76	0.29 ± 0.02
Blackcheek Tonguefish	33	32.21 ± 1.5	34.03 ± 1	0.38 ± 0.03
Gulf Pipefish	23	62.3 ± 4.4	64.96 ± 4.59	0.25 ± 0.05
Rough Silverside	20	46.7 ± 0.93	57.5 ± 1.21	1.43 ± 0.1
Code Goby	11	23.45 ± 1.48	28.64 ± 1.81	0.33 ± 0.05
Pinfish	7	72.29 ± 8.59	90.71 ± 11.49	14.3 ± 3.35
Chain Pipefish	6	70 ± 7.45	73 ± 7.93	0.3 ± 0.11
Scaled Sardine	5	47.4 ± 1.17	59.6 ± 1.29	2.26 ± 0.19
Speckled Seatrout	$\overline{4}$	38.75 ± 4.64	47.75 ± 6.02	1.14 ± 0.33
Atlantic Threadfin	$\overline{2}$	46.5 ± 3.5	56 ± 4	1.79 ± 0.31
Dwarf Seahorse	$\overline{2}$	$NA \pm NA$	18.5 ± 0.5	0.03 ± 0.02
Bay Anchovy	$\mathbf{1}$	$30 \pm NA$	$37 \pm NA$	$0.27 \pm NA$
Freckled Blenny	$\mathbf{1}$	$25 \pm NA$	$28 \pm NA$	$0.12 \pm NA$
Gulf Killifish	1	$19 \pm NA$	$24 \pm NA$	$0.14 \pm NA$
Silver Perch		$70 \pm NA$	$86 \pm NA$	$7.29 \pm NA$

Table 1.3 *Count and morphological metrics for fish collected, NA values indicate data not collected or not applicable for metric.*

Table 1.4 *Fish assemble metrics for 22 sites sampled*

Three GAMs were formulated to identify potential drivers of the fish assemblage metrics of density, species richness, and Shannon diversity. The first model with total number of fish as the response variable identified the wet biomass *R. maritima* of vegetation recovered from throw trap (χ 2= 6.992, p<0.05) as the only significant predictor variable (Table 1.5), and the second and third models identified no significant predictor variables for species richness and Shannon diversity (Tables 1.6 and 1.7).

 $*$ Value significant (p<0.05)

Table 1.5 *GAM output for potential drivers of site density*

 $*$ Value significant (p<0.05)

Table 1.6 *GAM output for potential drivers of site species richness*

Variable	χ2	
Depth (cm)	0.008	0.931
Wet biomass (g) of R . maritima recovered from throw trap	0.117	0.759

 $*$ Value significant (p<0.05)

Table 1.7 *GAM output for potential drivers of site Shannon diversity*

1.4 Discussion

The goal of this project was to describe the spatial distribution of reproductive *R. maritima* across the Chandeleur Islands and investigate differences in seagrass-associated fishes between reproductive and non-reproductive plants. To accomplish these goals, we conducted extensive seagrass surveys along the length of the Chandeleur Islands and used throw trapping to describe fish assemblages.

Ruppia maritma was found along the entire distribution of seagrass at North Chandeleur Island. Given the robustness of *R. maritima* to environmental stress, the absence of the species at only three of the sampled sites is likely linked to localized physical disturbance. The only other species present at the stations sampled was turtlegrass (*Thalassia testudinum*). Turtlegrass is a climax species that requires relatively stable environmental conditions and low levels of physical disturbance (Larkum et al., 2006). Turtlegrass occurred in centralized portions of the islands that likely represent areas with relatively more stable environmental and physical conditions.

Similar to turtlegrass, reproductive *R. maritima* plants were primarily restricted to the center of the island's back shelf. The complex reproductive shoots have reduced structural tissue and rely on support from the surrounding water (Kantrud, 1997), making them susceptible to physical disturbances. The central portion of the island represents an area with increased sheltering from wind driven wave action that dominates the system. The most northern and southern portions of the islands are characterized by reduced island relief and more exposed shallow waters (Kahn, 1986; Miselis & Plant, 2021). It is possible that *R. maritima* is reproductive at the northern and southern tips of North Chandeleur island, but that shoots cannot sustain the physical disturbance and are dislodged, suggesting that the environmental requirements for sexually reproductive *R. maritima* are stricter than those of non-reproductive plants.

Core samples showed distinct differences between reproductive and nonreproductive shoot densities. Shifts in shoot density have been described for *R. maritima* during the growth phase leading up to reproduction, where resources may be selectively partitioned to the reproductive shoots to promote vegetative growth, resulting in lowered survivorship of the non-reproductive shoots (Bigley & Harrison, 1986). Among the cores collected, less than one third (29.2%) showed measurable levels of epiphyte cover. The lack of epiphytes may be due to changes in bed characteristics as described above, where older non-reproductive shoot density is reduced when beds rapidly produce fresh reproductive vegetation, as well as due to the rapid growth of reproductive plants. The

rapid growth may not provide sufficient time for epiphytes to colonize a measurable amount.

R. maritima exhibits morphological plasticity across its range in distribution (Cho et al., 2009; Ito et al., 2013; Lopez-Calderon et al., 2010; Martínez-Garrido et al., 2017), and the current study represents the first known description of *R. maritima* reproductive shoot morphology at the Chandeleur Islands. Results from this study of increased surface area and leaf number for reproductive shoots relative to non-reproductive shoots are consistent with majority of descriptions for the species (Bigley & Harrison, 1986; Kantrud, 1991), but plasticity in reproductive plant morphology has been observed among habitat types (Richardson, 1983). In this study, the mean shoot length for reproductive shoots was 229.37 mm, which is much shorter than the maximum described shoot length of 2.5 m (Larkum et al., 2006). The observed difference may be linked to the presence of large physical disturbances that limit the growth of the fragile reproductive stems, differences in population structure, and/or may be due to a variety of other environmental drivers (Richardson, 1983).

This study describes patterns and potential drivers of habitat use by fish in reproductive and non-reproductive *R. maritima* and suggests that total plant biomass most influences density in *R. maritima* meadows. Similar relationships have been described for nekton communities of *R. maritima* in brackish ponds in Louisiana (Kanouse et al., 2006). The significant relationship between total plant biomass and proportion of *R. maritima* present implies that biomass can be a proxy for overall complexity. GAMs for species richness and Shannon diversity showed no predictor variable relationships were significant, further suggesting that the impacts of biomass and complexity are marginal

when compared to the overall influence of seagrass presence for shaping the fish community. This conclusion aligns with previous studies investigating relationships between seagrass presence and biomass and animal abundance, diversity, and richness (Heck et al., 2003; Strayer et al., 2003; Wyda et al., 2002). These results suggest that future studies to investigate the role of *R. maritima* in structuring fish communities in similar study systems may be better served to focus effort on robust measures of seagrass complexity (e.g. biomass) rather than more laborious measures of plant complexity such as cores and image analyses.

The lack of significant predictors among GAMs for species richness and Shannon diversity implies that other predictors are driving the relationships. One possible driver may be prey availability, which is linked to the function of seagrass as foraging grounds. Invertebrates were collected during the present study but were found to be outside the size range of those consumed by the fishes collected. Future studies designed to collect infauna and smaller invertebrates could help elucidate the relationship between prey availability and habitat use. Additionally, other environmental drivers such as salinity (Matheson et al., 1999), turbidity (Blaber & Blaber, 1980), and the availability of dissolved nutrients (Deegan et al., 2002) may impact the distribution of fishes within seagrass habitats. Although salinity was measured in the present study, there was little variation in the measured values which limited power in any subsequent analysis.

The fish collected within *R. maritima* meadows in this study were predominately small benthic species with individuals under 100-mm total length. The majority of individuals were adults of their respective species, with notable exceptions being spotted seatrout (*Cynoscion nebulosus*) and pinfish (*Lagodon rhomboides*) which were composed of young-of-year individuals (*FishBase*, 2022). The family Gobiidae made up the majority (52.7%) of all species collected. Gobiidae are benthic-associated fish and are known to live within seagrass meadows during all life stages where they feed on meiofaunal prey and use the structure provided by seagrasses as shelter from predators (Ara et al., 2010; Carle & Hastings, 1982). Although adult pinfish are known to be abundant in seagrass beds at the Chandeleur Islands (e.g., Hayes 2021), they were collected in low numbers during this study. This may be due to the life history characteristics of the species, where adults leave seagrass beds between May and October to spawn (Faletti et al., 2019), which could have led to a decrease in abundance within seagrass beds at the time of this study (September).

Interestingly, the fish collected in this study included no tropical species. Recent studies have reported the occurrence of tropical fishes in Chandeleur Islands seagrass meadows, indicating a northward movement of these species and tropicalization (Fodrie et al., 2010; Hayes, 2021). However, the previous studies collected relatively low abundances of tropical species (e.g. Lane Snapper *(Lutjanus synagris)* 0.056 per m² and Gag Grouper (*Mycteroperca microlepis*) 0.002 per m²; (Hayes, 2021)), sampled using trawls and benthic sleds which covered a greater area than the throw trap, and targeted different seagrass species (e.g. *Thalassia testudinum*, Hayes 2021). The absence of tropical species in our small-scale (1 m^2) samples indicates that, although they may be present in seagrass meadows at the Chandeleur Islands, tropical species are not likely abundant in *R. maritima* meadows.

One potential limitation of the current study is the throw trap sampling method which may underrepresent the abundance of highly mobile species (Freeman et al., 1984;

Kushlan, 1981) such as members of family Mugilidae. Despite this limitation, throw traps have been shown to have high rates of accuracy when describing fish assemblages (Jordan et al., 1997). Additionally, the removal of all vegetation within the throw trap contributed to clearing efficiency, as animals were recovered from the collected plant material during processing at GCRL.

1.5 Conclusion

R. maritima occurred along the entire latitudinal range of seagrass distribution at North Chandeleur Island. Despite the widespread distribution of non-reproductive plants, there was a central spatial trend of reproductive activity. Where reproductive plants occurred, their density was lower than that of non-reproductive plants, and reproductive shoots had significantly greater shoot lengths, number of leaves, and surface area when compared to non-reproductive plants. Non-reproductive plants showed significantly greater RSR, which indicates resource allocation to belowground biomass rather than aboveground biomass. These results of distinct spatial and morphological characteristics for reproductive *R. maritima* plants suggest that the ecosystem functions of *R. maritima* may change with the onset of reproduction.

Fish assemblages within *R. maritima* beds at the Chandeleur Islands are represented by an abundant group of small benthic species. The presence of morphologically complex reproductive *R. maritima* plants had little impact on the fish assemblages; rather, total biomass of seagrass was the primary drivers of habitat use. *R. maritima* has increased in cover by colonizing areas of disturbance along the islands and displacing climax species such as *T. testudinum*. Future studies should compare *R.*

maritima to other co-occurring seagrass species to better understand possible functional differences for associated fishes.

1.6 Project Significance

The Chandeleur Islands represent a highly dynamic system that supports the most diverse seagrass community in the region. The rise in occurrence of *R. maritima* within the system has been linked to large-scale disturbances that are increasing in frequency along the islands. This study described the spatial distribution and reproductive ecology of *R. maritima* at the Chandeleur Islands and is the first known description of reproductive morphology of this species for the area. *R. maritima* was present across most sites surveyed, often in monospecific meadows, suggesting that it has the ability to thrive in areas where other seagrass species may not. The occurrence of reproductive plants in high densities at the Chandeleur Islands also reinforces the predominance of this life history strategy for the species, and future research should investigate the role of sexual reproduction in the spread of *R. maritima* across the Chandeleur Islands. Results of this study also indicate that, despite the distinct morphologies between reproductive and non-reproductive plants, there were no discernible impacts on habitat use by fishes. Rather, the biggest driver was the more robust metric of total seagrass biomass.

Regardless of its reproductive status, *R. maritima* supported diverse and productive fish communities at the Chandeleur Islands. The recent decline in seagrass cover along the Chandeleur Islands has elicited attention by natural resources managers and in response to land and seagrass loss, a large-scale restoration project is in the early planning phase with a goal of preventing further decline. Managers seeking to restore seagrass along the Chandeleur Islands should view *R. maritima* as a species that hosts

diverse and abundant fish communities and should consider this species in restoration efforts to provide long-term resilience and stability of seagrass ecosystem function within the system.

	Area	Number	Aboveground	Belowground	Root to	Reprod-
Station	(mm ²)	of	Dried	Dried	Shoot	uctive
		Leaves	Biomass (g)	Biomass (g)	Ratio	
1897	294.973	4	0.009	0.005	0.584	$\mathbf N$
1897	585.090	3	0.017	0.014	0.811	${\bf N}$
1897	227.490	3	0.005	0.013	2.694	${\bf N}$
1897	366.621	$\overline{3}$	0.010	0.008	0.857	${\bf N}$
1897	489.886	$\overline{2}$	0.009	0.004	0.438	${\bf N}$
1897	4893.703	70	0.188	0.014	0.072	$\mathbf Y$
1897	1930.005	25	0.075	0.009	0.126	$\mathbf Y$
1897	1266.102	21	0.043	0.011	0.256	Y
1897	1442.344	19	0.060	0.004	0.067	Y
1897	1101.431	19	0.040	0.010	0.236	Y
C101	200.076	3	0.007	NA	NA	${\bf N}$
C101	268.251	3	0.009	NA	NA	${\bf N}$
C101	352.434	3	0.008	NA	NA	${\bf N}$
C101	308.179	3	0.008	NA	NA	${\bf N}$
C101	167.985	$\overline{2}$	0.004	NA	NA	${\bf N}$
C101	1610.557	22	0.104	NA	NA	Y
C113	1898.590	5	0.067	NA	NA	${\bf N}$
C113	819.093	$\overline{4}$	0.028	NA	NA	${\bf N}$
C113	989.616	$\overline{4}$	0.036	NA	NA	${\bf N}$
C113	959.831	$\overline{4}$	0.042	NA	NA	${\bf N}$
C113	533.139	3	0.023	NA	NA	${\bf N}$
C113	1949.759	48	0.102	NA	NA	Y
C113	1400.908	28	0.079	NA	NA	Y
C113	970.098	23	0.061	NA	NA	$\mathbf Y$
C113	886.280	20	0.055	NA	NA	Y
C113	759.661	20	0.044	NA	NA	Y
C ₁₁₅	271.917	5	0.007	NA	NA	$\mathbf N$
C ₁₁₅	258.675	$\overline{4}$	0.012	NA	NA	$\mathbf N$
C115	120.273	3	0.004	NA	NA	$\mathbf N$
C ₁₁₅	76.385	$\overline{2}$	0.003	NA	NA	N
C115	102.404	$\overline{2}$	0.004	NA	NA	N
C115	2568.365	50	0.130	NA	NA	Y
C115	1786.163	42	0.107	NA	NA	Y

Ratio, and Reproductive Status

Station	Length	Reproducti	
	(mm)	-ve	
C52	99.3560	${\bf N}$	
C52	210.4970	${\bf N}$	
C52	197.9930	${\bf N}$	
C52	100.9500	$\mathbf N$	
C52	101.9920	${\bf N}$	
C ₅₂	144.3140	Y	
C ₅₂	137.1010	Y	
C ₅₂	172.5170	Y	
C52	139.6640	Y	
C52	128.2180	Y	
C60	95.9500	${\bf N}$	
C60	97.9680	${\bf N}$	
C60	175.8340	$\mathbf N$	
C60	199.6340	$\mathbf N$	
C60	203.8590	N	
C60	208.7940	Y	
C60	306.0150	Y	
C60	270.2420	Y	
C60	165.9510	Y	
C60	192.9630	Y	
C68	220.2860	$\mathbf N$	
C68	224.0080	${\bf N}$	
C68	108.5630	$\mathbf N$	
C68	179.1310	${\bf N}$	
C68	277.0920	$\mathbf N$	
C68	286.5600	Y	
C68	233.5820	Y	
C68	385.8050	Y	
C68	336.2330	Y	
C68	186.6800	Y	
C76	82.4710	N	
C76	100.4860	N	
C76	75.9030	N	
C76	83.6530	N	
C76	88.4150	N	
C76	159.6720	Y	
C76	130.1560	Y	

APPENDIX C– Individual *R. maritima* Shoot Length and Reproductive Status

Individual Wet Weight

APPENDIX F– IACUC Approval Letter

REFERENCES

- Ambo-Rappe, R. (2016). Differences in Richness and Abundance of Species Assemblages in Tropical Seagrass Beds of Different Structural Complexity. *Journal of Environmental Science and Technology*, *9*(3), 246–256. https://doi.org/10.3923/jest.2016.246.256
- Ara, R., Arshad, A., Amin, S. M. N., Daud, S. K., Bujang, J. S., & Ghaffar, M. A. (2010). *Feeding habits of larval fishes of the family Gobiidae (Actinopterygii: Perciformes) in seagrass beds of Sungai Pulai estuary, Johor Strait, Malaysia*. 7.
- Bertelli, C. M., & Unsworth, R. K. F. (2014). Protecting the hand that feeds us: Seagrass (Zostera marina) serves as commercial juvenile fish habitat. *Marine Pollution Bulletin*, *83*(2), 425–429. https://doi.org/10.1016/j.marpolbul.2013.08.011
- Bethel, M., & Martinez, L. (2008). *Assessment of Current Seagrass Critical Habitat in Response to Dramatic Shoreline Change Resulting from the 2005 Hurricane Season for the Chandeleur Islands*. 12.
- Bigley, R. E., & Harrison, P. G. (1986). Shoot demography and morphology of Zostera japonica and Ruppia maritima from British Columbia, Canada. *Aquatic Botany*, *24*(1), 69–82. https://doi.org/10.1016/0304-3770(86)90118-X
- Blaber, S. J. M., & Blaber, T. G. (1980). Factors affecting the distribution of juvenile estuarine and inshore fish. *Journal of Fish Biology*, *17*(2), 143–162. https://doi.org/10.1111/j.1095-8649.1980.tb02749.x
- Camp, E. V., Gwinn, D. C., Lauretta, M. V., Pine, W. E., & Frazer, T. K. (2011). Use of Recovery Probabilities Can Improve Sampling Efficiency for Throw Traps in

Vegetated Habitats. *Transactions of the American Fisheries Society*, *140*(1), 164– 169. https://doi.org/10.1080/00028487.2011.558778

- Carle, K. J., & Hastings, P. A. (1982). Selection of Meiofaunal Prey by the Darter Goby, Gobionellus boleosoma (Gobiidae). *Estuaries*, *5*(4), 316–318. https://doi.org/10.2307/1351755
- Castillo-Rivera, M., Zavala-Hurtado, J. A., & Zárate, R. (2002). Exploration of spatial and temporal patterns of fish diversity and composition in a tropical estuarine system of Mexico. *Reviews in Fish Biology and Fisheries*, *12*, 167–177. https://doi.org/10.1023/A:1025051027676
- Cho, H. J., Biber, P., & Nica, C. (2009). *THE RISE OF RUPPIA IN SEAGRASS BEDS: CHANGES IN COASTAL ENVIRONMENT AND RESEARCH NEEDS*. 15.
- Collins, M., Sutherland, M., Bouwer, L., Cheong, S.-M., Combes, H. J. D., Roxy, M. K., Losada, I., McInnes, K., Ratter, B., Rivera-Arriaga, E., Susanto, R. D., Swingedouw, D., Tibig, L., Bakker, P., Eakin, C. M., Emanuel, K., Grose, M., Hemer, M., Jackson, L., … Turley, C. (2019). *Extremes, Abrupt Changes and Managing Risk. In: IPCC Special Report on the Ocean and Cryosphere in a Changing Climate*. 68.
- Cuadros, A., Cheminée, A., Thiriet, P., Moranta, J., Vidal, E., Sintes, J., Sagristá, N., & Cardona, L. (2017). The three-dimensional structure of Cymodocea nodosa meadows shapes juvenile fish assemblages at Fornells Bay (Minorca Island). *Regional Studies in Marine Science*, *14*, 93–101. https://doi.org/10.1016/j.rsma.2017.05.011
- Darnell, K. M., Carruthers, T. J. B., Biber, P., Georgiou, I. Y., Michot, T. C., & Boustany, R. G. (2017). Spatial and Temporal Patterns in Thalassia testudinum Leaf Tissue Nutrients at the Chandeleur Islands, Louisiana, USA. *Estuaries and Coasts*, *40*(5), 1288–1300. https://doi.org/10.1007/s12237-017-0229-y
- Darnell, K. M., Furman, B. T., Heck Jr., K. L., Byron, D., Reynolds, L., & Dunton, K. H. (2021). Seed Reserve Hot Spots for the Sub-Tropical Seagrass Halodule wrightii (Shoal Grass) in the Northern Gulf of Mexico. *Estuaries and Coasts*, *44*(2), 339– 351. https://doi.org/10.1007/s12237-020-00808-y
- Deegan, L. A., Wright, A., Ayvazian, S. G., Finn, J. T., Golden, H., Merson, R. R., & Harrison, J. (2002). Nitrogen loading alters seagrass ecosystem structure and support of higher trophic levels. *Aquatic Conservation: Marine and Freshwater Ecosystems*, *12*(2), 193–212. https://doi.org/10.1002/aqc.490
- Ellinwood, M. C. (2008). *Response of barrier island fish assemblages to impacts from multiple hurricanes: Assessing resilience of Chandeleur Island fish assemblages to hurricanes Ivan (2004) and Katrina (2005)*. 140.
- Faletti, M. E., Chacin, D. H., Peake, J. A., MacDonald, T. C., & Stallings, C. D. (2019). Population dynamics of Pinfish in the eastern Gulf of Mexico (1998-2016). *PLOS ONE*, *14*(8), e0221131. https://doi.org/10.1371/journal.pone.0221131

FishBase. (2022). https://www.fishbase.se/search.php

Fodrie, F. J., Heck, K. L., Powers, S. P., Graham, W. M., & Robinson, K. L. (2010). Climate-related, decadal-scale assemblage changes of seagrass-associated fishes in the northern Gulf of Mexico. *Global Change Biology*, *16*(1), 48–59. https://doi.org/10.1111/j.1365-2486.2009.01889.x

- Francis, T. (2013). *Fish Species Distribution in Seagrass Habitats of Chesapeake Bay are Structured by Abiotic and Bioti*. 12.
- Freeman, B. J., Greening, H. S., & Oliver, J. D. (1984). Comparison of Three Methods for Sampling Fishes and Macroinvertebrates in a Vegetated Freshwater Wetland. *Journal of Freshwater Ecology*, *2*(6), 603–609. https://doi.org/10.1080/02705060.1984.9664643
- Fujiwara, M., Martinez-Andrade, F., Wells, R. J. D., Fisher, M., Pawluk, M., & Livernois, M. C. (2019). Climate-related factors cause changes in the diversity of fish and invertebrates in subtropical coast of the Gulf of Mexico. *Communications Biology*, *2*(1), 1–9. https://doi.org/10.1038/s42003-019-0650-9
- Handley, L., & Lockwood, C. (2020). *2020 Seagrass Status and Trends Update*. https://cnlworld-my.sharepoint.com/personal/handleyn_cnlworld_org/_layouts
- Hartog, C. den, & Kuo, J. (2006). Taxonomy and Biogeography of Seagrasses. In A. W. D. LARKUM, R. J. ORTH, & C. M. DUARTE (Eds.), *SEAGRASSES: BIOLOGY, ECOLOGYAND CONSERVATION* (pp. 1–23). Springer Netherlands. https://doi.org/10.1007/978-1-4020-2983-7_1
- Hayes, C. (2021). *Patterns of Habitat Use and Trophic Structure in Turtle Grass (Thalassia testudinum)-Dominated Systems Across the Northern Gulf of Mexico*. 283.
- Heck, K., Hays, G., & Orth, R. (2003). Critical evaluation of the nursery role hypothesis for seagrass meadows. *Marine Ecology Progress Series*, *253*, 123–136. https://doi.org/10.3354/meps253123
- Hemminga, M. A., & Duarte, C. M. (2000). *Seagrass Ecology*. Cambridge University Press.
- Hitchcock, J. K., Courtenay, S. C., Coffin, M. R. S., Pater, C. C., & van den Heuvel, M. R. (2017). Eelgrass Bed Structure, Leaf Nutrient, and Leaf Isotope Responses to Natural and Anthropogenic Gradients in Estuaries of the Southern Gulf of St. Lawrence, Canada. *Estuaries and Coasts*, *40*(6), 1653–1665. https://doi.org/10.1007/s12237-017-0243-0
- Hori, M., Suzuki, T., Monthum, Y., Srisombat, T., Tanaka, Y., Nakaoka, M., & Mukai, H. (2009). High seagrass diversity and canopy-height increase associated fish diversity and abundance. *Marine Biology*, *156*(7), 1447–1458. https://doi.org/10.1007/s00227-009-1184-3
- Horinouchi, M. (2007). Review of the effects of within-patch scale structural complexity on seagrass fishes. *Journal of Experimental Marine Biology and Ecology*, *350*(1– 2), 111–129. https://doi.org/10.1016/j.jembe.2007.06.015
- Hovel, K., Fonseca, M., Myer, D., Kenworthy, W., & Whitfield, P. (2002). Effects of seagrass landscape structure, structural complexity and hydrodynamic regime on macrofaunal densities in North Carolina seagrass beds. *Marine Ecology Progress Series*, *243*, 11–24. https://doi.org/10.3354/meps243011
- Hyndes, G. A., Kendrick, A. J., MacArthur, L. D., & Stewart, E. (2003). Differences in the species- and size-composition of fish assemblages in three distinct seagrass habitats with differing plant and meadow structure. *Marine Biology*, *142*(6), 1195–1206. https://doi.org/10.1007/s00227-003-1010-2
- Ito, Y., Ohi-Toma, T., Murata, J., & Tanaka, N. (2013). Comprehensive phylogenetic analyses of the Ruppia maritima complex focusing on taxa from the Mediterranean. *Journal of Plant Research*, *126*(6), 753–762. https://doi.org/10.1007/s10265-013-0570-6
- Jordan, F., Coyne, S., & Trexler, J. C. (1997). Sampling Fishes in Vegetated Habitats: Effects of Habitat Structure on Sampling Characteristics of the 1-m2 Throw Trap. *Transactions of the American Fisheries Society*, *126*(6), 1012–1020. https://doi.org/10.1577/1548-8659(1997)126<1012:SFIVHE>2.3.CO;2
- Kahn, J. H. (1986). Geomorphic Recovery of the Chandeleur Islands, Louisiana, after a Major Hurricane. *Journal of Coastal Research*, *2*(3), 337–344.
- Kanouse, S., La Peyre, M., & Nyman, J. (2006). Nekton use of Ruppia maritima and nonvegetated bottom habitat types within brackish marsh ponds. *Marine Ecology Progress Series*, *327*, 61–69. https://doi.org/10.3354/meps327061
- Kantrud, H. A. (1991). *Wigeongrass (Ruppia Maritima L.): A Literature Review*. United States Department of the Interior, Fish and Wildlife Service.
- Kantrud, H. A. (1997). *Wigeongrass (Ruppia maritima): A Literature Review.* https://agris.fao.org/agris-search/search.do?recordID=AV20120104488
- Kenworthy, W. J., Cosentino-Manning, N., Handley, L., Wild, M., & Rouhani, S. (2017). Seagrass response following exposure to Deepwater Horizon oil in the Chandeleur Islands, Louisiana (USA). *Marine Ecology Progress Series*, *576*, 145–161. https://doi.org/10.3354/meps11983
- Kilminster, K., McMahon, K., Waycott, M., Kendrick, G. A., Scanes, P., McKenzie, L., O'Brien, K. R., Lyons, M., Ferguson, A., Maxwell, P., Glasby, T., & Udy, J.

(2015). Unravelling complexity in seagrass systems for management: Australia as a microcosm. *Science of The Total Environment*, *534*, 97–109. https://doi.org/10.1016/j.scitotenv.2015.04.061

- Kushlan, J. (1981). Sampling Characteristics of Enclosure Fish Traps. *Transactions of The American Fisheries Society - TRANS AMER FISH SOC*, *110*, 557–562. https://doi.org/10.1577/1548-8659(1981)110<557:SCOEFT>2.0.CO;2
- Larkum, A. W. D., Orth, R. J., & Duarte, C. M. (Eds.). (2006). *Seagrasses: Biology, ecology, and conservation*. Springer.

Lopez-Calderon, J., Riosmena-Rodríguez, R., Rodríguez-Baron, J. M., Carrión-Cortez, J., Torre, J., Meling-López, A., Hinojosa-Arango, G., Hernández-Carmona, G., & García-Hernández, J. (2010). Outstanding appearance of Ruppia maritima along Baja California Sur, México and its influence in trophic networks. *Marine Biodiversity*, *40*(4), 293–300. https://doi.org/10.1007/s12526-010-0050-3

- Mahoney, R. D., Kenworthy, M. D., Geyer, J. K., Hovel, K. A., & Joel Fodrie, F. (2018). Distribution and relative predation risk of nekton reveal complex edge effects within temperate seagrass habitat. *Journal of Experimental Marine Biology and Ecology*, *503*, 52–59. https://doi.org/10.1016/j.jembe.2018.02.004
- Malea, P., Kevrekidis, T., & Mogias, A. (2004). Annual versus perennial growth cycle in Ruppia maritima L.: Temporal variation in population characteristics in Mediterranean lagoons (Monolimni and Drana Lagoons, Northern Aegean Sea). *Botanica Marina - BOT MAR*, *47*, 357–366. https://doi.org/10.1515/BOT.2004.052

Martínez-Garrido, J., Creed, J. C., Martins, S., Almada, C. H., & Serrão, E. A. (2017). First record of Ruppia maritima in West Africa supported by morphological description and phylogenetic classification. *Botanica Marina*, *60*(5), 583–589. https://doi.org/10.1515/bot-2016-0128

Matheson, R. E., Camp, D. K., Sogard, S. M., & Bjorgo, K. A. (1999). Changes in Seagrass-Associated Fish and Crustacean Communities on Florida Bay Mud Banks: The Effects of Recent Ecosystem Changes? *Estuaries*, *22*(2), 534. https://doi.org/10.2307/1353216

- McDevitt-Irwin, J., Iacarella, J., & Baum, J. (2016). Reassessing the nursery role of seagrass habitats from temperate to tropical regions: A meta-analysis. *Marine Ecology Progress Series*, *557*, 133–143. https://doi.org/10.3354/meps11848
- McKenzie, L. J., Yaakub, S. M., Tan, R., Seymour, J., & Yoshida, R. L. (2016). Seagrass habitats of Singapore: Environmental drivers and key processes. *RAFFLES BULLETIN OF ZOOLOGY*, 18.
- Middleton, M. J., Bell, J. D., Burchmore, J. J., Pollard, D. A., & Pease, B. C. (1984). Structural differences in the fish communities of Zostera capricorni and Posidonia australis seagrass meadows in Botany Bay, New South Wales. *Aquatic Botany*, *18*(1–2), 89–109. https://doi.org/10.1016/0304-3770(84)90082-2

Miselis, J. L., & Plant, N. G. (2021). Satellite-Derived Barrier Response and Recovery Following Natural and Anthropogenic Perturbations, Northern Chandeleur Islands, Louisiana. *Remote Sensing*, *13*(18), 3779. https://doi.org/10.3390/rs13183779

Moore, L. J., Patsch, K., List, J. H., & Williams, S. J. (2014). The potential for sea-levelrise-induced barrier island loss: Insights from the Chandeleur Islands, Louisiana, USA. *Marine Geology*, *355*, 244–259.

https://doi.org/10.1016/j.margeo.2014.05.022

- Nagelkerken, I., Roberts, C., van der Velde, G., Dorenbosch, M., van Riel, M., Cocheret de la Morinière, E., & Nienhuis, P. (2002). How important are mangroves and seagrass beds for coral-reef fish? The nursery hypothesis tested on an island scale. *Marine Ecology Progress Series*, *244*, 299–305. https://doi.org/10.3354/meps244299
- Nakaoka, M. (2005). Plant–animal interactions in seagrass beds: Ongoing and future challenges for understanding population and community dynamics. *Population Ecology*, *47*(3), 167–177. https://doi.org/10.1007/s10144-005-0226-z
- Nordlund, L. M., Unsworth, R. K. F., Gullström, M., & Cullen‐Unsworth, L. C. (2018). Global significance of seagrass fishery activity. *Fish and Fisheries*, *19*(3), 399– 412. https://doi.org/10.1111/faf.12259
- Olds, A. D., Connolly, R. M., Pitt, K. A., & Maxwell, P. S. (2012). Primacy of seascape connectivity effects in structuring coral reef fish assemblages. *Marine Ecology Progress Series*, *462*, 191–203. https://doi.org/10.3354/meps09849

Olesen, B., Marba, N., Duarte, C. M., Savela, R. S., & Fortes, M. D. (2004). Recolonization dynamics in a mixed seagrass meadow: The role of clonal versus sexual processes. *Estuaries*, *27*(5), 770–780. https://doi.org/10.1007/BF02912039

- Orth, R. J., & Moore, K. A. (1988). Distribution of Zostera marina L. and Ruppia maritima L. sensu lato along depth gradients in the lower Chesapeake Bay, U.S.A. *Aquatic Botany*, *32*(3), 291–305. https://doi.org/10.1016/0304-3770(88)90122-2
- Pham, L. T., Biber, P. D., & Carter, G. A. (2014). Seagrasses in the Mississippi and Chandeleur Sounds and Problems Associated with Decadal-Scale Change Detection. *Gulf of Mexico Science*, *32*(1). https://doi.org/10.18785/goms.3201.03

Poirrier, M., & Handley, L. (2007). *Chandeleur Islands*.

- Pulich, W. M. (1985). *SEASONAL GROWTH DYNAMICS OF RUPPIA MARITIMA L. s.1, AND HALODULE WRIGHTII ASCHERS. IN SOUTHERN TEXAS AND EVALUATION OF SEDIMENT FERTILITY STATUS*. 14.
- Radloff, P., Hobson, C., Whisenant, A., & Bronson Warren, J. (2013). *Statewide seagrass monitoring protocol development—Phase 2 final report*.
- Reyes, E., & Merino, M. (1991). Diel Dissolved Oxygen Dynamics and Eutrophication in a Shallow, Well-Mixed Tropical Lagoon (Cancun, Mexico). *Estuaries*, *14*(4), 372–381. https://doi.org/10.2307/1352262
- Richardson, F. (1983). *Variation, Adaptation and Reproductive Biology in Ruppia Maritima L. Populations from New Hampshire Coastal and Estuarine Tidal Marshes* [Ph.D., University of New Hampshire]. https://www.proquest.com/docview/303170779/abstract/1DFCCCF60C204E5BP $Q/1$
- Rotherham, D., & West, R. J. (2002). Do different seagrass species support distinct fish communities in south-eastern Australia?: SEAGRASS FISH COMMUNITIES.

Fisheries Management and Ecology, *9*(4), 235–248.

https://doi.org/10.1046/j.1365-2400.2002.00301.x

- Skilleter, G., Loneragan, N., Olds, A., Zharikov, Y., & Cameron, B. (2017). Connectivity between seagrass and mangroves influences nekton assemblages using nearshore habitats. *Marine Ecology Progress Series*, *573*, 25–43. https://doi.org/10.3354/meps12159
- Staveley, T. A. B., Hernvall, P., Stjärnkvist, N., van der Meijs, F., Wikström, S. A., & Gullström, M. (2020). Exploring seagrass fish assemblages in relation to the habitat patch mosaic in the brackish Baltic Sea. *Marine Biodiversity*, *50*(1), 1. https://doi.org/10.1007/s12526-019-01025-y
- Staveley, T. A. B., Perry, D., Lindborg, R., & Gullström, M. (2017). Seascape structure and complexity influence temperate seagrass fish assemblage composition. *Ecography*, *40*(8), 936–946. https://doi.org/10.1111/ecog.02745
- Strayer, D. L., Lutz, C., Malcom, H. M., Munger, K., & Shaw, W. H. (2003). Invertebrate communities associated with a native (Vallisneria americana) and an alien (Trapa natans) macrophyte in a large river. *Freshwater Biology*, *48*(11), 1938–1949. https://doi.org/10.1046/j.1365-2427.2003.01142.x
- Vaslet, A., Phillips, D. L., France, C., Feller, I. C., & Baldwin, C. C. (2012). The relative importance of mangroves and seagrass beds as feeding areas for resident and transient fishes among different mangrove habitats in Florida and Belize: Evidence from dietary and stable-isotope analyses. *Journal of Experimental Marine Biology and Ecology*, *434–435*, 81–93. https://doi.org/10.1016/j.jembe.2012.07.024
- Williams, J. A., Holt, G. J., Robillard, M. M. R., Holt, S. A., Hensgen, G., & Stunz, G. W. (2016). Seagrass fragmentation impacts recruitment dynamics of estuarinedependent fish. *Journal of Experimental Marine Biology and Ecology*, *479*, 97– 105. https://doi.org/10.1016/j.jembe.2016.03.008
- Wyda, J., Deegan, L., Hughes, J., & Weaver, M. (2002). The response of fishes to submerged aquatic vegetation complexity in two ecoregions of the Mid-Atlantic Bight: Buzzards Bay and Chesapeake Bay. *Estuaries and Coasts*, *25*, 86–100. https://doi.org/10.1007/BF02696052